

Asian Journal of Research in Infectious Diseases

8(2): 23-29, 2021; Article no.AJRID.74315 ISSN: 2582-3221

Pseudomonas aeruginosa Infection and Antibiotic Sensitivity Pattern Isolated from a Tertiary Care Hospital in Dhaka City

Mousumi Karmaker^{1*}, Md. Abul Khair¹, Una Jessica Sarker¹, Rabeya Nahar Ferdous¹, Sa'dia Tasnim², Mohammad Moniruzzaman² and Shah Md. Zahurul Haque Asna¹

¹Department of Microbiology, Bangladesh University of Health Sciences, Bangladesh. ²Department of Immunology, Bangladesh University of Health Sciences, Bangladesh.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJRID/2021/v8i230233 <u>Editor(s):</u> (1) Dr. Win Myint Oo, SEGi University, Malaysia. <u>Reviewers:</u> (1) Bawane Rashmi subhash, MUHS, India. (2) Aline Pereira Pedrosa, Federal University of Rio de Janeiro, Brazil. (3) Mohammed Ansar Qureshi, College of Medicine. Najran University, Saudi Arabia. Complete Peer review History: <u>https://www.sdiarticle4.com/review-history/74315</u>

Original Research Article

Received 19 July 2021 Accepted 29 September 2021 Published 08 October 2021

ABSTRACT

Pseudomonas aeruginosa is one of the most widespread gram-negative microorganisms identified in the clinical samples and most common causes of hospital acquired infection. *P. aeruginosa* is affecting both indoor and outdoor patients throughout the world. Due to frequent mutation in *P. aeruginosa* highly resistant strain developed rapidly. The aim of the study to determine the prevalence of *P. aeruginosa* species in different samples isolated from a Tertiary care Hospital as well as determination their diverse antibiotic resistance pattern. This cross-sectional study was carried out to determine in-vitro resistance pattern of *P. aeruginosa* isolates to common antimicrobial agents by disc diffusion method. Various clinical samples were collected from Bangladesh Health Sciences Hospital (BIHS) General Hospital, Dhaka. This research was carried out in the Department of Microbiology of Bangladesh University of Health Sciences (BUHS). Isolation, identification and antibiogram were performed for *P. aeruginosa* following standard microbiological laboratory procedure. A total of 218 *P. aeruginosa* were isolated from 3062 different

*Corresponding author: E-mail: moniruzzaman@buhs.ac.bd;

clinical specimens which are statistically significant (p<0.0001). Among the highest number of *P. aeruginosa* were isolated from outdoor patients 140 compare to Indoor patients which are significantly higher (p <0.013). In this study Male (68.3%) are more vulnerable to *P. aeruginosa* infection compare to females (31.7%) which is also statistically significant. Young people (less than 35 years) were more susceptible to *P. aeruginosa* infection which is also statistically significant (p< 0.01). The highest number of *P. aeruginosa* was isolated from wound (43.12%), followed by pus (40.33%), sputum (8.71%) urine (7.80%). The maximum number of *P. aeruginosa in various samples* was resistant to aztreonam and co-tromoxazole followed by cephalosporins, aminoglycosides, carbapenems. The most sensitive antibiotic was colistin of followed by gentamycin and tetracycline. To control the spread of resistant bacteria, it is disparagingly vital to have stringent antibiotic guidelines. The antibiotic susceptibility pattern of *P. aeruginosa* requires to be continuously monitored in specialized clinical units and the results readily made available to the clinicians to minimize the resistance.

Keywords: Pseudomonas aeruginosa' AmpC beta-lactamase; muller-hinton agar media; antimicrobial susceptibility testing; colistin.

1. INTRODUCTION

aeruginosa is found in the hospital Ρ. environment and therefore it is the most common causative agent of nosocomial infections. The bacterium possesses a wide range of secretion systems, which secretes numerous proteins relevant to the pathogenesis of clinical strains [1]. P. aeruginosa is found throughout the body, most commonly in the urinary tract, respiratory tract, blood, and wounds [2]. P. aeruginosa is a type of bacterium that can develop resistance to rather rapidly antibiotics over several generations. This resistance present in some strains makes P. aeruginosa very difficult to treat infected hosts such as a human or other animal [3]. Production of a variety of beta-lactamases, outer membrane permeability and combinations of multiple mechanisms of resistance, the gramnegative bacteria are becoming resistant to beta lactam such cephalosporins, drugs as monobactams and carbapenems [4]. Some strains of P. aeruginosaproduce AmpC betalactamase enzyme and this enzyme make this strain MDR (multi drug resistance) bacterium. As P. aeruginosa is a commensal in healthy people the frequency of commensalism risesgradually with the duration of hospital stay [5]. P. aeruginosa is frequently isolated as an opportunistic pathogen in occurring often infection of hospitalized patients and has been isolated from a number of sites in the hospital environment [6]. P.aeruginosa is the most important, resistant and dangerous organism infecting the burn patient [7]. It is the fifth pathogen among common hospital microorganisms and causes 10% of all hospital acquired infection [8]. In Bangladesh, it ranks third and causes a wide range of infection [9].

Recently this bacterium has become increasingly resistant to various antimicrobial agent [10]. Drugs resistant *P.aeruginosa* isolated has emerged and continues to escalate rapidly [11]. Over the period of time we observed an increase in the number of *P. aeruginosa* among laboratory isolates so we decide to carry out a cross-sectional study to see infections caused by *P. aeruginosa* and the susceptibility pattern of the organism isolated from a different clinical specimens at the department of microbiology, Bangladesh University of Health Science.

2. MATERIALS AND METHODS

cross-sectional laboratory-based This retrospective study was carried out in the Department of Microbiology, BUHS from 1st July 2017to 31st August 2018. A total of 218 P.aeruginosa were isolated from 3062 clinical specimens including wound swab, pus, urine, sputum. To avoid contamination, wounds and pus samples were thoroughly cleaned with sterile normal saline followed by gentle rubbing of the wound site with 70% alcohol before swabbing samples. Urine and sputum samples were also collected by maintaining an appropriate procedure. The isolates were identified by colony characters, Gram's staining and different biochemical test were performed; Triple sugar iron, citrate, Motility, Indole, Urea and the test for P.aeruginosa Confirmatory was performed Oxidase positive test (In case of blood, pus wound swab and urine sample). The antibiotic susceptibility patterns of *P.aeruginosa* isolates were analyzed by clinically used common antibiotics. Antibiotic Susceptibility test was carried out by disc diffusion method (Kirby-Bauer) in Muller-Hinton agar media according to

the CLSI. M100 Performance Standards for Antimicrobial Susceptibility Testing, 30th edition. The results of the susceptibility test were interpreted into Sensitive, Intermediate and resistant [12].

3. RESULTS

The cross-sectional study was conducted on 3062 patients. Among 3062 samples, 218 *P.aeruginosa* were isolated and the proportion was 7.1 (95% Cl 6.2 to 8.0) (p<0.0001). Where most of the *P.aeruginosa* (64.2%) was isolated from outdoor patients and 35.8% were from indoor patients, which is significant. Maximum (86.2%) patients were estimated above 35 years, and the rest of (13.8%) were below 35 years. 68.3% *P. aeruginosa* were predominantly isolated from male patients and female patients 31.7% (Table 1).

Among 218 isolates of *P. aeruginosa*, 94 were isolated wound samples (43.12%) which is highest in number following, 88(40.33%) from pus, 19 (8.71%) from sputum and 17(7.80%) from urine (Table 2).

The resistance rate of *P.aeruginosa* isolated from different specimens to different antimicrobials wide-ranging from 0.01% to 100%. P. aeruginosa isolated from pus, wound and urine sample were found 100% resistant to aztreonam and cotrimoxazole. Isolates of pus samples found (82.9%) (89.7%), (84.1%), resistance to ciprofloxacin, doxycycline and cefixime antibiotics respectively. P. aeruginosaof pus and wound samples were found (84.1%) and (64.8%) resistance to carbapenem groups of antibiotics respectively. However, pus isolates were found (94.3%) sensitive to Colistin. P. aeruginosa isolated from wound samples were estimated (95.7%) and (92.6%) resistance to ceftazidime and doxycycline respectively. P. aeruginosa isolated from sputum samples were assessed (89.5%) susceptible to imipenam, gentamicin, amikacin and tigecycline. However, Ρ. aeruginosa isolated from sputum samples were found (100%) resistance to aztreonam. P. aeruginosa isolated from urine samples were found (88.2 %) sensitive to gentamicin and amikacin and (88.2%), (94.1%) and (100%) resistance to ceftazidime, doxycycline, cotrimoxazole and aztreonam respectively. On the other hand, P. aeruginosa isolated from urine and sputum samples were found (82.4%) and (89.5 %) sensitive to carbapenem groups of antibiotics respectively. Where colistin were found more effective drug against all kind of P. aeruginosa infection. P. aeruginosa isolates of wound sample were found (71.3 %) sensitive to colistin. P. aeruginosa isolated from sputum samples were found to (99%) sensitive to colistin (Table 3).

4. DISCUSSION

P. aeruginosa emerged as a common pathogen for nosocomial infections. It is one of the important causes of illness among indoor and outdoor patients in the hospital. The increasing resistance rate of P. aeruginosa in hospital infections is due to its incomplete and indiscriminate use of antibiotics. In the present study, it is manifest that there is diversity in the resistant pattern of isolates of P. aeruginosa from different clinical samples. In our study, the maximum of P. aeruginosa were identified from patients who were aged above 35 years which is 188 (86.2%) compared to below 35 years which is 30 (13.8%). However, the study of The National Nosocomial Infections Surveillance System showed that aged between 21-40 years (41.2%) were more susceptible to P. aeruginosa infection [13]. Where male patients were more vulnerable to P. aeruginosa infection 149 (68.3%) comparison to female 69 (31.7%). In 2004 Friedland et al. showed a prevalence of P.aeruginosa species in outdoor patients was 70% and indoor patients were 30% [14]. In our study, we found the same parameter where outdoor patients (64.2%) prevalence is higher in reference with indoor patients (35.8%) with an increasing rate.

Variables		Number	Percentage	P value
Patient group	Indoor patient	78	35.8%	< 0.013
	Outdoor patient	140	64.2%	
Gender	Male	149	68.3%	< 0.01
	Female	69	31.7%	
Age group	Below 35	188	86.2%	< 0.01
	Above 35	30	13.8%	

Type of sample	No. of sample (n)	No of <i>P.aeruginosa</i> isolated	Percentages (%)
Urine	2116	17	7.80%
Pus	455	88	40.33%
Wound	306	94	43.12%
Sputum	185	19	8.71%
Total	3062	218	100%

Table 2.	Percentages of P	. aeruginosa	isolated fr	om both	indoor	and out	door	patients
different clinical samples (n=218)								

Antimicrobials	Resistance Proportion of Wound <i>P. aeruginosa</i> (n=94)	Resistance Proportion of Urine <i>P. aeruginosa</i> (n=17)	Resistance Proportion of Pus <i>P.</i> aeruginosa(n=88)	Resistance Proportion of Sputum <i>P.</i> aeruginosa(n=19)
Imepenam	64.8%	17.6%	84.1%	10.5%
Ciprofloxacin	81.0%	35.3%	84.1%	15.8%
Ceftazidime	95.7%	88.2%	86.3%	68.4%
Doxycycline	92.6%	94.1%	89.7%	73.7%
Gentamicin	79.8%	11.8%	76.1%	10.5%
Cefixime	89.4%	94.1%	82.9%	63.2%
Amikacin	76.6%	11.8%	73.9%	10.5%
Co-trimoxazole	100%	100%	100%	63.2%
Aztreonam	100%	100%	100%	100%
Tetracycline	36.2%	23.5%	23.9%	10.5%
Colistin	28.7%	5.9%	5.7%	0.01%

Maximum clinical isolates of *P.aeruginosa* were isolated from the wound (43.12%), followed by pus (40.33%), urine 17 (7.80%). These results are in line with studies of Houge MM et al. where they estimated (pus/ wound 38.38% and urine 28.28%) [15]. Similarly, Mousumi et al. in found the prevalence of Pseudomonas species in diabetic foot infected patients where pus samples were 22% and in case of non-diabetic foot infected patients where pus samples were 29% [16]. Though, in the study results of Teteeng et al. contradict our results they estimated, 19% P. aeruginosa from wound samples, and they identified the highest number of staphylococcus aureus (36%) from wound samples, these data contrast to our study [17]. In this study in case of sputum sample, only 8.71 % P. aeruginosa were isolated, similarly, Abdelraouf et al. identified 7.0% P. aeruginosa in sputum samples [18]. In this study, P.aeruginosa isolated from wound samples and pus were found more resistant in all commercially available antibiotic compare to P. aeruginosa isolated from urine and sputum samples. Most of the P. aeruginosa species isolated from all samples were found resistance to co-trimoxazole and aztreonam. On the other

hand, Abdelraouf et al. found P.aeruginosa highly resistant to amoxicillin and cephalexin [18]. In our study, P. aeruginosa isolated from pus samples were found 100% resistance to cotrimoxazole and aztreonam, 86.3% doxycycline, 82.9% cefixime, and 84.1% resistance to both imipenem and ciprofloxacin, 76.1% gentamicin, 73.9% amikacin. In 2015 Mousumi et al. estimated that only 33% P. aeruginosa were resistant to carbapenem groups of antibiotic which is now 100% [16]. Which indicates carbapenem resistant is increasing day by day. Though only 5.7% P. aeruginosa pus samples were found resistance to colistin. Teteeng et al. found only 8% ciprofloxacin resistance P. wound samples. But aeruginosa from comparable to our study Siddiqua et al. showed highest resistance (91% to 96%) to cotrimoxazole and cefuroxime P. aeruginosa isolated from wound samples [19]. P. aeruginosa isolated from urine samples were found 100% resistance to azitronam and co-trimoxazole as well as 94.1% to doxycycline and cefixime. 17.6% resistance to imipenem. Alike our study Ali H et al. found almost all of the P. aeruginosa isolates from urine samples were highly resistant to doxycycline but sensitive to amikacin (100%). norfloxacin (86%) ciprofloxacin (83%) and tobramycin (83%) [20]. Though, Fatima et al. estimated the highest resistance Р to aeruginosa isolate with piperacillin/ tazobactam and cefipime i.e. 42% and 40% respectively. Imipenem was found to be most effective antibiotic against P aeruginosa (76% sensitivity) amikacin resistance was continuously but increasing [21]. In our study, we also found imipenem and colistin also found effective drug against P aeruginosa isolated from sputum samples. Colistin (99.6% sensitive) proved to be one of the most effective drugs for routine use among the P. aeruginosa strains investigated in this study. Colistin therapy was at least as effective and as safe as beta-lactam antibiotic or a guinolone in the treatment of infections caused by multidrug-resistant P. aeruginosa [22].

5. CONCLUSION

In summary, P. aeruginosa strains were more commonly isolated from pus and wound samples. Carbapenem resistance P. aeruginosa strains are increasing day by day and our study also demonstrated that colistin is highly useful as a preferred and effective drug for all kinds of P. aeruginosa infection. Though resistance to colistin of pus sample is lower in comparison with other antibiotics, but it is really an issue of concern to found resistance to colistin antibiotic. The superiority of colistin as anti-pseudomonal therapy needs to be further verified through larger prospective clinical trials. To prevent the spread of resistant bacteria, it is critically important to have strict antibiotic guidelines while investigation programs for multidrug-resistant organisms and infection control procedures need to be executed. It is compulsory that the antibiotic sensitivity pattern of highly resistant bacterial pathogens like P. aeruginosa in particular clinical units to be continuously supervised and the results readily made accessible to the physician to control the morbidity.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by

the producing company rather it was funded by personal efforts of the authors.

CONSENT

We take consent from every patient during data and sample collection.

ETHICAL APPROVAL

This research was approved by the Ethical committee of Bangladesh University of Health Sciences.

ACKNOWLEDGEMENT

No financial support has been contributed to the development of this research. We are very thankful to the laboratory of the Department of Microbiology of BIHS General Hospital for their assistance.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Altopark U, Erol S, Akcay MN, Celebi F, Kadanali A. et al. Time related changes of antimicrobial resistance patterns and predominant bacterial profiles of burn wounds and bodyfloraof burned patients. Burns. 2004; 30(7):660-4. DOI: 10.1016/j.burns.2004.03.005
- Rashid A, Chowdhury A, Rahman SHZ, Begum SA, Muazzam N, et at. Infections by *Pseudomonas aeruginosa* and antibiotic resistance pattern of the isolates from Dhaka medical college hospital. BJMM. 2007;01(02):48-51. DOI:

https://doi.org/10.3329/bjmm.v1i2.21508

- Collee JK, Miles RS. Tests for identification of bacteria. In: Collee JG, Duguid JP, Faser AK, Mermoin BP, et al. Mackie & MacCartney practical Medical Microbiology, 13thed. New York; USA: Churchill Livingstone. 1998;141-160.
- 4. DrenkardE. Antimicrobial resistant of *Pseudomonas aeruginosa* biofilms. Microbes Infect. 2003;5(13):1213-1219. DOI: 10.1016/j.micinf.2003.08.009.
- 5. Emori TG, Gaynes RP. An overview of nosocomial infection, including the role of

the microbiology laboratory. Clin Microbiol Rev. 1993;6(4):428-42. DOI: 10.1128/CMR.6.4.428

6. Karimi H, Kashani PP, Ghanaatpisheh Frequency of *Pseudomonas aeruginosa* serotypes in burn wound infections and their resistance to antibiotics. Burns. 2002; 28(4):340-345.

DOI: 10.1016/s0305-4179(02)00024-4

- 7. Fridkin SK, Steward CD, Edwards JR, Pryor ER, McGowanJr JE. Archibald LK, Gaynes RP, Tenover FC.et al.Surveillance of Antimicrobial Use and antimicrobial Resistance in United States Hospitals. Clin Infect Dis.1999;29(2):245-52. DOI: 10.1086/520193
- Gailiene G, Pavilonis A, Kareiviene V. The peculiarities of *Pseudomonas aeruginosa* resistance to antibiotics and prevalence of serogroups. Medicina (Kaunas). 2007; 43(1):36-42. PMID: 17297282
- Kawser NM, Khan NK, Rahman ASM, Mondol EA. Study on nosocomial infections and antibiotic resistance pattern of the isolates in ICU patients. BAFMJ. 2002;2(7):9-13.
- 10. Neu,H.C.The role of *Pseudomonas aeruginosa* in infections. J Antimicrob Chemother. 1983;11(4:)1-13. DOI: 10.1093/jac/11.suppl b.1
- 11. Shahcheragi F, Feizabadi MM, Yamin V, Abiri R, Abedian Z.et al.Serovar determination, drug resistance patterns and plasmid profiles of *Pseudomonas aeruginosa* isolated from burn patients at two hospitals of Tehran (IRAN). Burns. 2003;29(6): 547-51.

DOI: 10.1016/s0305-4179(03)00142-6.

- Julia A. Kiehlbauch, George E. Hannett, Max Salfinger, Wendy Archinal, Catherine Monserrat, Cynthia Carlyn, et al. Use of the national committee for clinical laboratory standards guidelines for disk diffusion susceptibility testing in new york state laboratories. J Clin Microbiol. 2000;38(9):3341–3348. DOI: 10.1128/JCM.38.9.3341-3348.2000
- 13. Jones RN, Kirby JT, Beach ML, Biedenbach DJ, Pfaller MA, et al. Geogra phic variations in activity of broadspectrum beta-lactams against Pseudomonas aeruginosa: Summary of worldwide SENTRY Antimicrobial the Program (1997-2000). Surveillance Diagn Microbiol Infect Dis. 2002;43(3):239-243.

DOI: 10.1016/s0732-8893(02)00390-5.

14. Friedland I, Gallagher G, King T, Woods GL.et al.Antimicrobial susceptibility patterns in Pseudomonas aeruginosa: Data from a multicenter Intensive Care Unit Surveillance Study (ISS) in the United States. J Chemother. 2004;16(5): 437-41.

DOI: 10.1179/joc.2004.16.5.437

 Hoque MM, Ahmad M, Khisa S, Uddin MN, Jesmine R. Antibiotic Resistance Pattern in Pseudomonas Aeruginosa Isolated from Different Clinical Specimens. JAFMC Bangladesh. 2015;11(1):45–49. DOI:

https://doi.org/10.3329/jafmc.v11i1.30669

 Karmaker M, Sanyal SK, Sultana M, Hossain MA. Association of bacteria in diabetic and non-diabetic foot infection – An investigation in patients from Bangladesh. J. Infect Public Health. 2016;9(3):267-27.

DOI: 10.1016/j.jiph.2015.10.011

- 17. Tateeng YM, Enweani IB. Okogun GRA, Nwobu GO, Okodua M, Okpala HO,Agba MI,et al. Prevalence of *Pseudomonas aeruginosa* in post operative wound infections at the University College Hospital Ibadan. Environmental Health & Human Development. 2002;3(2). Available:https://www.researchgate.net/pu blication/282012435
- 18. Abdelraouf A. Elmanama Najah M Elaiwa Farid H, Abu-Elamreen Abed El-kader Y, El-Ottol, et al. *Pseudomonas aeruginosa* distribution in clinical sample and their antibiogram from Al-Shifa Hospital, Gaza, PNA. Annals of Alquds Medicine; 2006;2(1427-2006):37-45. Available:https://www.researchgate.net/pu

blication/230707321

- Siddiqua M, Alam AN, Akter S, Ferdousi RS, et al.Antibiotic resistance pattern in *Pseudomonas aeruginosa* isolated from a private Medical College Hospital. KYAMC Journal. 2018;9(1)12-19. DOI: 10.3329/kyamcj.v9i1.36617
- Salih HA, Abdulbary M, Abdulrida AS. Susceptibility of Pseudomonas aeruginosa isolated from urine to some antibiotics.AL-Qadisiya Journal of Veterinary Medicine Sciences (QIVMS). 2011;10:203-7. DOI:

https://doi.org/10.29079/vol10iss2art158

21. Fatima A, Naqvi SB, Khaliq SA ,Perveen S, Jabeen S, et al. Antimicrobial susceptibility pattern of clinical isolates of

Pseudomonas aeruginosa isolated from patients of lower respiratorytract infections. Springerplus. 2012;1(1):70. DOI: 10.1186/2193-1801-1-70

22. Hachem RY, Chemaly RF, Ahmar CA, Jiang Y, Boktour MR, Rjaili GA, Bodey GP, Raad II, et al. Colistin is effective in treatment of infections caused by multidrug-resistant *Pseudomonas aeruginosa* in Cancer Patients. Antimicrobial Agents and Chemotherapy. 2007;51(6):1905–11. DOI: 10.1128/AAC.01015-06

© 2021 Karmaker et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

> Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle4.com/review-history/74315